

Creating connections between biotechnology and industrial sustainability

August 25 to 28, 2024 Costão do Santinho Resort, Florianópolis, SC, Brazil

INDUSTRIAL MICROBIOLOGY: PROSPECTING AND APPLIED MOLECULAR BIOLOGY

# SCREENING OF CLOSTRIDIUM STRAINS PRODUCING BIOMOLECULES FROM THE PERSPECTIVE OF BIOREFINERIES

Pamela Baptista Ferreira<sup>1</sup>, Vanessa Kristine de Oliveira Schmidt<sup>2</sup>, Maikon Kelbert<sup>3</sup> & Débora de Oliveira<sup>3\*</sup>

<sup>1</sup> Chemical Engineering/Technological Center/Department of Chemical Engineering and Food Engineering/Federal University of Santa Catarina,

Florianópolis, Brazil. \*pamelabapferreira@gmail.com vanessa.schmidt@posgrad.ufsc.br maikonkelbert@gmail.com debora.oliveira@ufsc.br

## ABSTRACT

The production of biomolecules such as hydrogen and volatile fatty acids (VFAs) from lignocellulosic materials of agro-industrial waste is crucial both for converting waste into high-value products and for mitigating waste disposal issues. Hydrogen stands out as an efficient and clean energy alternative, while VFAs are crucial for the production of chemicals, fuels, and polymers. *Clostridium* strains are recognized for their capacity to produce these compounds via ABE fermentation. However, the industrial implementation of these bioprocesses still faces challenges in scalability and optimization, highlighting the importance of prospecting and evaluating new strains capable of producing these compounds for future applications. This study identified *Clostridium* strains with high potential for biohydrogen production, reaching a maximum production of  $6.15 \pm 1.15\%$  hydrogen through fermentation. Additionally, the production of butyric acid and butanol was observed through volatile compound analysis.

Keywords: Clostridium. Biohydrogen. Fermentation. Prospection. VFA.

## **1 INTRODUCTION**

Short-chain volatile fatty acids (VFAs), such as acetic acid and butyric acid, constitute relevant raw materials for various industrial sectors, including food, pharmaceutical, chemical, and biofuel industries (SUN; ZHANG; LOH, 2021). Hydrogen (H<sub>2</sub>), in turn, is widely recognized as a promising alternative energy source due to its environmentally friendly nature and high energy yield (122 kJ/g) (Rambabu et al., 2020). Although various technologies have been developed for the production of these compounds, the biotechnological method has garnered interest, mainly due to the use of lignocellulosic material (LCM) derived from agro-industrial residues. This method is particularly notable for its approach of integrated bioprocesses that employ innovative microorganisms (Rambabu et al., 2020).

The bacterial genus *Clostridium* is recognized for its ability to generate various bioproducts through ABE fermentation (Acetone-Butanol-Ethanol). Among these bioproducts, volatile fatty acids (VFAs) (e.g., butyric and acetic acid) and biohydrogen (bioH<sub>2</sub>) stand out, which can be produced via the acidogenic route. However, achieving industrial-scale production of these compounds with high yields and productivity through biotechnological pathways remains a significant challenge.

Recently, researchers have turned their attention to the prospecting of new strains with the potential for producing these biocompounds for possible large-scale implementation. According to Fonseca *et al.* (2016)., it was possible to isolate a new H<sub>2</sub>-producing bacterial culture from anaerobic sludge. The isolated culture produced the highest amount of H2 at 35 °C and an initial pH of 7, utilizing various carbon sources, including glucose, galactose, and mannose from algal biomass.

Although some studies have identified *Clostridium* strains that produce VFAs and bioH<sub>2</sub> (JIANG et al., 2018), comparative research between these strains is lacking, especially in the context of a biorefinery utilizing lignocellulosic material (LCM). Therefore, the objective of the study is to prospect new strains of Clostridium capable of efficiently synthesizing bioproducts of high industrial value, such as VFAs and bioH<sub>2</sub>.

## 2 MATERIAL & METHODS

The anaerobic granular sludge was taken from an anaerobic reactor from a food industry in the Santa Catarina state. The sludge (volatile solids:  $42.73 \pm 0.44$  g/L) was pre-treated by decreasing the pH to 3.0 before plating. The acidification was performed by adding the sludge sample (5 mL) to 0.1 M citrate phosphate buffer at pH 3.0 (35 mL) and incubating for 12 h at 37 °C with an agitation of 150 rpm.

After pre-treatment, 100 µL of sludge were diluted in 900 µL NaCl solution (0.9% w/v). Subsequently, 100 µL of the diluted sample were plated on sterile Petri dishes covered with 25 mL Differential Reinforced Clostridial Medium (DRCM) to culture *Clostridium* species at pH 7.0. Petri dishes were incubated in an anaerobic jar at 37\_°C until colonies appeared. To ensure colony isolation, the process was repeated three times. The colonies were resuspended in saline solution, and 1.0 mL was inoculated into serum

bottles containing 50 mL of RCM medium. Nitrogen gas was bubbled into the bottles before sealing with a rubber stopper and aluminum seal and incubated at 37°C for 72 h. After this period, the headspace gas composition was analyzed by gas chromatography. With the liquid portion, we conducted the analysis of VFAs (with gas chromatography-mass spectroscopy). The organisms producing VFA-H2 were then separated for identification. The methodology used was adapted as described by Fonseca *et al.* (2016).

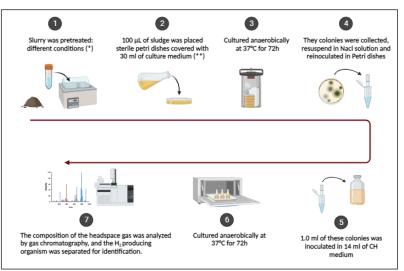
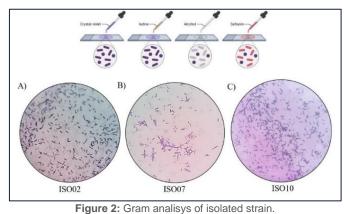


Figure 1: Description of the methodology used.

### **3 RESULTS & DISCUSSION**

1

The acid pretreatment of the sludge favored the isolation of three strains producing  $H_2$  and VFAs, named ISO02, ISO07, and ISO10. The DRCM revealed the presence of *Clostridium* by darkening of the medium due to sulfite reduction, common in *Clostridium spp*.



Microscopy images at 100x magnification show a positive result for the Gram test and the rod-shaped morphology of the isolated bacteria. The samples are: (A) ISO02, (B) ISO07, and (C) ISO10.

Preliminary results of gas chromatography confirmed H<sub>2</sub> ( $5.57\pm0.82$ ,  $6.15\pm1.15$ , and  $5.62\pm0.5\%$ ) and CO<sub>2</sub> ( $30.40\pm0.91$ ,  $20.23\pm1.06$ , and  $23.23\pm0.75\%$ ) production by the strains (Table 1).

Table 1: Quantification of hydrogen and carbon dioxide produced by the isolated stains by gas chromatografy.

Samples	Hydrogen (% mol/ mol)	Carbon Dioxide (% mol/ mol)
ISO02	5.57 ± 0.82	30.40 ± 0.91
ISO07	6.15 ± 1.15	20.23 ± 1.06
ISO10	$5.62 \pm 0.5$	23.23 ± 0.75

VFA analysis identified products such as butyric acid. Additionally, the presence of alcohols such as butanol was noted in Table 2.

				ISO 02	ISO 07	ISO10
Compound name	Formula	MW (g/mol)	RT	Average Peak area (%)	Average Peak area (%)	Average Peak area (%)
Alcohols						
Ethanol	C <sub>2</sub> H <sub>6</sub> O	46,07	1,98	1,98		0,35
1-Butanol	C <sub>4</sub> H <sub>10</sub> O	74,12	3,45	2,85	0,38	0,25
3-Thietanol	C <sub>3</sub> H <sub>6</sub> OS	90,14	3,96	10,17		
1-Hexanol	C <sub>6</sub> H <sub>14</sub> O	102,17	7,37	5,16		
1-Butanol, 3-methyl-	C <sub>5</sub> H <sub>12</sub> O	88,15	4,60	1,11	2,88	1,62
1-Nonanol	C <sub>9</sub> H <sub>20</sub> O	144,25	8,81	1,34		
1-Octanol	C <sub>8</sub> H <sub>18</sub> O	130,23	11,61	1,68		
Alkane						
Cyclotetrasiloxane, octamethyl-	C <sub>8</sub> H <sub>24</sub> O <sub>4</sub> Si <sub>4</sub>	296,62	14,59	0,49	2,19	2,45
Benzene, (1-hexylheptyl)-	C19H32	260,46	23,79	0,10		
Chlorinated compound						
Phenol, 2-chloro-4-(1,1-dimethylpropyl)-	C11H15CIO	198,69	18,03	0,15		13,00
Ester						
Phytol, acetate	C22H42O2	338,57	23,88	0,23		
Di-sec-butyl phthalate	C16H22O4	278,34	24,35	0,13		
Acetic acid, hexyl ester	C <sub>8</sub> H <sub>16</sub> O <sub>2</sub>	144,21	10,43	0,14		
Propanoic acid, 2-methyl-, 3-methylbutyl ester	C <sub>9</sub> H <sub>18</sub> O <sub>2</sub>	158,24	11,34			15,45
Butanoic acid, 3-methyl-, 3-methylbutyl ester	C10H20O2	172,26	12,29		0,24	3,81
Butanoic acid, 3-methylbutyl ester	C <sub>9</sub> H <sub>18</sub> O <sub>2</sub>	158,24	11,31	8,55	2,43	0,32
Organic Acid						
Butanoic acid	C <sub>4</sub> H <sub>8</sub> O <sub>2</sub>	88,11	6,70	3,78	2,34	
Butanoic acid, 3-methyl-	C <sub>5</sub> H <sub>10</sub> O <sub>2</sub>	102,32	7,62		0,83	

Table 2: The main volatile organic compounds produced by ISO02, ISO07, ISO10.

Soluble metabolites analysis revealed the presence of various compounds, as shown in Table 2. Changes in pH significantly affect the byproducts of fermentative processes. A pH below 6.0 favors the production of butyric and acetic acids, while a higher pH results in acetic acid and ethanol. In *Clostridium* species, the transition from pH 7 to 4.5 shifts the phase from acidogenic to solventogenic. The accumulation of organic acids and bioH<sub>2</sub> reduces pH, leading to the solventogenic phase and decreasing the biosynthesis of these compounds.

#### **4 CONCLUSION**

The results confirm the isolation of *Clostridium* strains with the potential for biomolecules production, as well as demonstrating the production of bioH<sub>2</sub> and VFAs by these isolated strains.

#### REFERENCES

- SUN, J.; ZHANG, L.; LOH, K. C. Review and perspectives of enhanced volatile fatty acids production from acidogenic fermentation of lignocellulosic biomass wastes. Bioresources and Bioprocessing, v. 8, n. 1, p. 1–21, 2 ago. 2021.
- K. Rambabu, P.-L. Show, G. Bharath, F. Banat, M. Naushad, and J.-S. Chang. Enhanced biohydrogen production from date seeds by *clostridium thermocellum* atcc 27405. International Journal of Hydrogen Energy, 45(42):22271–22280, 2020. ISSN 0360-3199.
- JIANG, L. et al. Butyric acid: Applications and recent advances in its bioproduction. Biotechnology Advances, v. 36, n. 8, p. 2101–2117, 2018.
- AFSAH-HEJRI, L. et al. Identification of volatile organic compounds (VOCs) by SPME-GC-MS to detect Aspergillus flavus infection in pistachios. Food Control, v. 154, n. May, p. 110033, 2023.
- FONSECA, B. C. et al. Fermentative production of H2 from different galactose concentrations by the new isolate *Clostridium beijerinckii* Br21. *International Journal of Hydrogen Energy*, 41(46), 21109–21120. Dec. 14. 2016.
- KANDIA B., RONDAGS E., FRAMBOISIER X., MAUVIEL G., DUFOUR A., GUEDON E, Diauxic growth of *Clostridium acetobutylicum* ATCC 824 when grown on mixtures of glucose and cellobiose. *AMB Express*, v. 8, n. 1, 2018.
- STOKLOSA, R. J. et al. Butyric Acid Generation by *Clostridium tyrobutyricum* from Low-Moisture Anhydrous Ammonia (LMAA) Pretreated Sweet Sorghum Bagasse. *Applied Biochemistry and Biotechnology*, v. 193, n. 3, p. 761–776, 2021.

#### ACKNOWLEDGEMENTS

I would like to express my gratitude to my professors and colleagues at LiEB, where I conducted most of my experiments. I also thank CNPq, CAPES and SHV Energy/Supergasbras for the scholarship and funding.